

PRELIMINARY NOTES

BBA 41 153

Effect of uncouplers on the transport of anions in rat-liver mitochondria

The inner mitochondrial membrane appears to contain specific translocators mediating the transport of anionic substrates^{1,2}. In rat-liver mitochondria, exchange-diffusion reactions involving α -oxoglutarate and dicarboxylate^{3,4}, tricarboxylate and malate (ref. 4 and A. J. MEYER, unpublished observations), and P_i and dicarboxylate⁵ have been directly demonstrated. The stoichiometry of these reactions is one to one³⁻⁵. Evidence has also been obtained⁵ that the translocation of phosphate, but not that of malate (contrast ref. 6), can be directly coupled to an OH^- counterflux. The specificity of these reactions, and their sensitivity to inhibitors, indicate that they are mediated by separate translocators^{5,7}.

Uncouplers inhibit the accumulation of anionic substrates by mitochondria⁸⁻¹⁰, the inhibition being competitive with respect to the anions¹¹. VAN DAM AND SLATER¹² proposed that uncoupler anions compete with substrate anions for entry into the mitochondria, the uncoupler anion being transported *via* a non-specific anion translocator (see, however, refs. 11, 13). In view of the fact that the translocators are

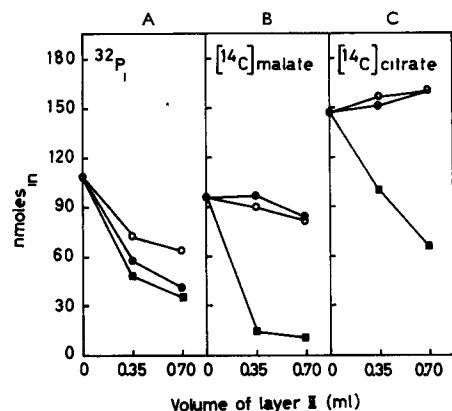


Fig. 1. Effect of dicoumarol on the efflux of anionic substrates from rat-liver mitochondria. Mitochondria (Expt. A, 9.2 mg protein, Expts. B and C, 7 mg protein) were preincubated in 1 ml of 150 mM sucrose, 20 mM Tris-Cl (pH 7.4), 0.5 mM EDTA, 1 mM $MgCl_2$, 1.4 μ g rotenone, 0.34 μ g antimycin and 10 μ g oligomycin. After 1 min, 1 mM $^{32}P_i$ ($9 \cdot 10^5$ counts/min; Expt. A), 1 mM [^{14}C]malate ($5 \cdot 10^5$ counts/min; Expt. B) or 1 mM [^{14}C]citrate ($1 \cdot 10^6$ counts/min; Expt. C) were added. The mitochondria in 0.35 ml of the preincubation mixture were centrifuged¹⁴ through a washing layer containing the same components as the preincubation mixture (except labelled substrate), through a second incubation layer, and finally into $HClO_4$. Layer II contained the same components as the preincubation mixture (except labelled substrate), with no additions (○—○), 40 μ M dicoumarol (●—●); 5 mM unlabelled P_i (Expt. A, ■—■) or 5 mM unlabelled malate (Expts. B and C, ■—■). The time in Layer II was about 15 sec when the volume was 0.35 ml. The amount of labelled substrate remaining in the matrix space was calculated from the radioactivity in the $HClO_4$ extract. The values refer to the original amount of mitochondrial protein.

specific, that only phosphate exchanges with hydroxyl, and that different translocators can be linked through the anions shared, the problem arises of the specificity of the action of uncouplers on the different transport systems. An insight into this has been obtained by examining the effect of uncouplers on the efflux of various anions involved in the exchanges, both in the absence and in the presence of the counter anions.

Mitochondria were preloaded with a labelled substrate anion in the presence of oligomycin, antimycin, and rotenone, and then centrifuged¹⁴ through a washing layer, through a second layer containing the substances whose effect on the efflux of the anion was to be studied, and finally into HClO₄. The amount of anion remaining in the matrix space of the mitochondria was determined. In Fig. 1, the efflux of ³²P_i, [¹⁴C]malate and [¹⁴C]citrate is shown. In the controls (no additions to the second incubation layer), a substantial efflux of P_i occurred, but in contrast, no significant efflux of malate or citrate was observed. The ³²P_i efflux was increased by unlabelled P_i. Unlabelled malate promoted a rapid and extensive efflux of both [¹⁴C]malate and [¹⁴C]citrate. Dicoumarol significantly increased the efflux of ³²P_i, but had practically no effect on the matrix level of either [¹⁴C]malate or [¹⁴C]citrate.

The data in Table I show that dicoumarol or 2,4-dinitrophenol promoted the efflux of P_i, but not that of citrate or α -oxoglutarate (*cf.* Fig. 1). Malate promoted

TABLE I

EFFECT OF UNCOUPLERS AND INHIBITORS ON THE EFFLUX OF ANIONIC SUBSTRATES FROM RAT-LIVER MITOCHONDRIA

Experimental procedure as in Fig. 1. Expt 1: 7 mg mitochondrial protein, preincubation with 1 mM ³²P_i (1·10⁶ counts/min) or 1 mM [¹⁴C]citrate (7·10⁵ counts/min). Expt 2: 6.8 mg mitochondrial protein; preincubation with 1 mM ³²P_i or 1 mM α -oxoglutarate. Expt 3: 8.4 mg mitochondrial protein, preincubation with 1 mM ³²P_i. Expt 4: 11.2 mg mitochondrial protein; preincubation with 1 mM ³²P_i; 300 μ M mersalyl added 30 sec after addition of ³²P_i. Centrifugation filtration began 3 min after addition of labelled substrate or mersalyl. Layer II contained where indicated 40 μ M dicoumarol, 100 μ M 2,4-dinitrophenol, 5 mM malate, and 5 mM butyl malonate. The time in Layer II was about 15 sec. α -Oxoglutarate was estimated enzymically.

Expt No	Additions	Amount (nmoles) in matrix of		
		³² P _i	[¹⁴ C]Citrate	α -Oxoglutarate
1	None	47	124	
	Dicoumarol	39	127	
	Dinitrophenol	40	130	
	Malate	32	63	
2	None	66		35
	Dicoumarol	54		33
	Malate	44		2
3	None	66		
	Dinitrophenol	50		
	Butyl malonate	80		
	Dinitrophenol + butyl malonate	50		
4	None	77		
	Dinitrophenol	63		
	Mersalyl	95		
	Dinitrophenol + mersalyl	106		

the efflux of P_i , citrate and α -oxoglutarate. Butyl malonate and mersalyl inhibited the efflux of P_i that occurred in the absence of an added anion. Mersalyl, but not butyl malonate, prevented the 2,4-dinitrophenol-induced efflux of P_i , indicating that it is mediated only by the phosphate-hydroxyl translocator⁷

TABLE II

EFFECT OF DICOUMAROL ON v_{max} OF MALATE EFFLUX FROM RAT-LIVER MITOCHONDRIA AND THE K_m FOR THE COUNTER ANION

Using the experimental procedure described in Fig 1, mitochondria (80 mg in Expts 1 and 3, 70 mg in Expts 2 and 4) preloaded with [^{14}C]malate were centrifuged through the second incubation layer containing 0.1 mM counter anion with or without 100 μM dicoumarol. The counter anion-induced efflux of [^{14}C]malate, *i.e.* the difference in the amount of [^{14}C]malate remaining in the matrix after passage of the mitochondria through Layer II in the absence and presence of counter anion, was determined. These values were used to construct Lineweaver-Burk plots, from which the values for K_m and v_{max} were obtained.

Expt No	Counter anion	v_{max} (nmoles/15 sec)		K_m (μM)	
		Control	+ Dicoumarol	Control	+ Dicoumarol
1	P_i	45	48	131	400
2	Malate	67	67	23	40
3	Citrate	57	51	102	217
4	α -Oxoglutarate	51	40	56	84

The effect of uncouplers on the counter anion-driven efflux of [^{14}C]malate was examined kinetically (Table II). Dicoumarol inhibited the exchange-diffusion reactions involving malate, the inhibition being purely competitive in the case of the malate-malate and malate-phosphate antiports, and partly competitive in other cases. A similar inhibition of the malate-phosphate and malate- α -oxoglutarate exchanges was obtained with 2,4-dinitrophenol. However, it had no effect on the malate-citrate or malate-malate exchanges. Possibly, the malate-phosphate, malate- α -oxoglutarate and malate-tricarboxylate translocators must all be inhibited in order to inhibit the malate-malate exchange; this appears to be the case with dicoumarol, but not with 2,4-dinitrophenol. VAN DAM AND KRAAYENHOF¹⁵ have shown that 2,4-dinitrophenol inhibits the accumulation of citrate by freshly prepared mitochondria. However, the uptake of citrate under their conditions involves not only an exchange of citrate with intramitochondrial citrate and malate, but also a recycling of the endogenous malate and phosphate *via* the malate-phosphate translocator. Furthermore, the stimulation by 2,4-dinitrophenol of the efflux of P_i will contribute to the inhibition of [^{14}C]citrate uptake.

Our results suggest that the mechanism proposed by KRAAYENHOF AND VAN DAM¹¹ to explain the inhibition by uncouplers of the uptake of anions can be applicable in the case of the phosphate-hydroxyl antiporter. In the other cases studied here, the inhibition must be due at least in part, to an interaction of the uncoupler with the translocator that does not appear to involve transport of the uncoupler anion (see also ref. 6). In other experiments (to be reported elsewhere) we have found that the dinitrophenol-induced efflux of $^{32}P_i$ can be accompanied by accumulation of [^{14}C]dinitrophenol by the mitochondria, both processes being mersalyl-sensitive. Further investigation is required to elucidate the exact mechanism of this coupling.

between phosphate efflux and uncoupler accumulation. The mechanisms proposed by MITCHELL⁶ and by VAN DAM^{11,12,15} could both, in principle, accommodate our findings.

This investigation was supported by a grant from the Consiglio Nazionale delle Ricerche, Italy, and by grants from the Life Insurance Medical Research Fund and from the Netherlands Foundation for Chemical Research (S.O.N.) with financial assistance from the Netherlands Organization for the Advancement of Pure Research (Z.W.O.) A.J.M. is grateful to the Consiglio Nazionale delle Ricerche for a Research Fellowship. The authors wish to thank J. L. Howland for a gift of [¹⁴C]dinitrophenol.

Department of Biochemistry,
University of Bari,
Bari (Italy) and
Laboratory of Biochemistry*,
University of Amsterdam,
Amsterdam (The Netherlands)

N. E. LOFRUMENTO
A. J. MEYER
J. M. TAGER
S. PAPA
E. QUAGLIARIELLO

- 1 J. B. CHAPPELL, *Brit. Med. Bull.*, 24 (1968) 150
- 2 E. J. DE HAAN AND J. M. TAGER, *Abstr. 3rd Meeting Federation European Biochem. Soc.*, Warsaw, 1966, Academic Press and Polish Scientific Publishers, London and Warsaw, 1966, p. 159
- 3 S. PAPA, R. D'ALOYA, A. J. MEYER, J. M. TAGER AND E. QUAGLIARIELLO, in S. PAPA, J. M. TAGER, E. QUAGLIARIELLO AND E. C. SLATER, *The Energy Level and Metabolic Control in Mitochondria*, Adriatica Editrice, Bari, Italy, 1969, p. 159
- 4 E. QUAGLIARIELLO, S. PAPA, A. J. MEYER AND J. M. TAGER, in L. ERNST AND Z. DRAHOTA, *Mitochondria: Structure and Function*, Academic Press, London, in the press
- 5 S. PAPA, N. E. LOFRUMENTO, M. LOGLISCI AND E. QUAGLIARIELLO, *Biochim. Biophys. Acta*, 189 (1969) 311
- 6 P. MITCHELL, *Chemiosmotic Coupling and Energy Transduction*, Glynn Research Ltd., Bodmin, Kent, 1968
- 7 A. J. MEYER AND J. M. TAGER, *Biochim. Biophys. Acta*, 189 (1969) 136
- 8 J. L. GAMBLE JR., *J. Biol. Chem.*, 240 (1965) 2668
- 9 E. J. HARRIS, K. VAN DAM AND B. C. PRESSMAN, *Nature*, 213 (1967) 1126
- 10 E. QUAGLIARIELLO AND F. PALMIERI, *European J. Biochem.*, 4 (1968) 20
- 11 R. KRAAYENHOF AND K. VAN DAM, *Biochim. Biophys. Acta*, 172 (1969) 189
- 12 K. VAN DAM AND E. C. SLATER, *Proc. Natl. Acad. Sci. U.S.A.*, 58 (1967) 2015
- 13 R. D. VELDSEMA-CURRIE AND E. C. SLATER, *Biochim. Biophys. Acta*, 162 (1968) 310
- 14 E. PFAFF, Ph.D. Thesis, Marburg, 1965
- 15 K. VAN DAM AND R. KRAAYENHOF, in S. PAPA, J. M. TAGER, E. QUAGLIARIELLO AND E. C. SLATER, *The Energy Level and Metabolic Control in Mitochondria*, Adriatica Editrice, Bari, Italy, 1969, p. 299

Received August 29th, 1969

* Postal address: Plantage Muidergracht 12, Amsterdam-C, The Netherlands